

Age-related lysosomal dysfunction: an unrecognized roadblock for cobalamin trafficking?

Hua Zhao · Ulf T. Brunk · Brett Garner

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Abstract Vitamin-B₁₂ is a generic term for corrinoid compounds that exhibit the biological activity of cyanocobalamin and are collectively referred to as cobalamins. Methylcobalamin and 5-deoxyadenosylcobalamin are the active cobalamins in human metabolism. Cobalamin plays a crucial role in the maintenance of homocysteine and methylmalonyl-CoA homeostasis and is required for erythrocyte formation and DNA synthesis. Data from human and animal studies indicate that cobalamin deficiency impairs neuronal function; a process that is thought to contribute to age-related cognitive decline and dementia. Cobalamin deficiency also results in dysfunction of the peripheral nervous system; among other disorders. Although there is a detailed understanding of the biochemical pathways that are perturbed in cobalamin deficiency, the mechanisms underlying age-related dyshomeostasis in such pathways remain to be addressed. Because cobalamin utilization is dependent on its efficient transit through lysosomes, and mounting evidence indicates that lysosomal function deteriorates in aging long-lived post-mitotic cells such as neurons, in the present article we

review published data that supports the proposition that impaired lysosomal processing of cobalamin may play a significant role in age-related (neuro) degenerative diseases.

Keywords Aging · Lysosomes · Lipofuscin · Vitamin-B12 · Neurodegeneration

Introduction

Vitamin-B₁₂ is a generic term for corrinoid compounds that exhibit the biological activity of cyanocobalamin and are collectively referred to as cobalamins. Methylcobalamin (MeCbl) and 5-deoxyadenosylcobalamin (AdoCbl) are the cobalamins that are active in human metabolism. Cobalamin (Cbl) is required for erythrocyte formation and DNA synthesis and plays a crucial role in maintenance of neurological function. Data from human and animal studies indicate that cbl deficiency impairs neuronal function; a process that is thought to contribute to age-related cognitive decline and dementia including Alzheimer's disease [14, 37, 51, 71]. Cbl deficiency also results in dysfunction of the peripheral nervous system [46, 62]; among other disorders [30]. Although there is a detailed understanding of the biochemical pathways that are perturbed in cbl deficient states (see below), several questions remain regarding why such biochemical/metabolic perturbations increase with age. Prompted by the fact that cbl utilization is critically dependent on its efficient transit through the lysosomal compartment [10, 21], and the evidence that lysosomal function deteriorates in aging long-lived post-mitotic cells such as neurons [13, 19, 22, 55, 59, 61], in the present report we review the available literature in order to explore the idea that suboptimal lysosomal processing of cbl may play a role in age-related loss of neurological function.

H. Zhao · B. Garner
Illawarra Health and Medical Research Institute,
University of Wollongong, Wollongong,
NSW 2522, Australia
e-mail: hz739@uowmail.edu.au

H. Zhao · B. Garner (✉)
School of Biological Sciences, University of Wollongong,
Wollongong, NSW 2522, Australia
e-mail: brettg@uow.edu.au

U. T. Brunk
Department of Pharmacology,
Linköping University, 581 85 Linköping, Sweden
e-mail: ulf.brunk@liu.se

Cbl absorption

Cbl (Fig. 1) is a water-soluble vitamin present in limited amounts in the diet, primarily in meat and dairy products. Clinically, total serum cbl concentrations are typically examined for deficiency status with serum cbl levels <150 pM and/or hematological anomalies treated as cbl deficiency. However, low serum cbl concentrations do not accurately reflect intracellular cbl status and thus serum homocysteine (Hcy) and methylmalonic acid (MMA) levels (>13 μ M and >0.4 μ M, respectively) are more reliable indicators of cbl deficiency [18]. These compounds accumulate in cbl deficiency as a result of reduced activities of two key enzymes, methionine synthase (MS) and methylmalonyl-CoA mutase (MMCM), that require cbl as a cofactor (discussed further below). Due to inadequate dietary intake and poor absorption, approximately 6% of the western population over the age of 60 has low serum cbl levels; with the prevalence of deficiency increasing with age [2].

Pernicious anemia accounts for approximately 15–25% of cbl deficiency. This disorder is characterized by a lack of intrinsic factor (IF), which limits the body's capacity to absorb cbl from dietary sources [18]. Pernicious anemia is an autoimmune disease that results in the destruction of gastric parietal cells which results in a lack of IF production. The resulting atrophic gastritis eventually leads to megaloblastic anemia and neurological disorders if left untreated. The majority of cbl deficiency (about 60–70%), however, is due to food-Cbl malabsorption, a problem of particular concern in the elderly [15]. This syndrome is characterized by the inability of the body to release cbl from food or intestinal transport proteins. These factors

clearly make a major contribution to functional cbl deficiency.

Dietary cbl undergoes carrier-mediated transport, first binding to salivary haptocorrin, from where it is released to IF in the duodenum. The intestinal absorption of IF-Cbl is mediated by cubilin receptors that are tethered to the surface of ileal mucosal cells via the amnionless protein [20, 25]. The IF-Cbl complex is sorted through lysosomes, the IF is degraded by lysosomal hydrolases, and cbl is freed and transported out of the lysosomal compartment. Cbl is then secreted via the basolateral membrane into the portal vein and transported to tissues as a complex with transcobalamin (TC) via the systemic circulation [43]. Plasma TC-Cbl is then internalized by the transcobalamin receptor (TCR) expressed on the cell surface (discussed further below). Plasma haptocorrin also binds cbl where it is thought to act as a store since it is not endocytosed by the TCR. It is noteworthy that intravenous cbl rapidly passes the blood–brain-barrier in humans, although the receptor(s) involved have not been definitively identified [63].

Cbl intracellular trafficking—the importance of lysosomes

Once the TC-Cbl complex is endocytosed by cells in the body, it is targeted to the lysosome where TC is degraded and the TCR is recycled to the cell surface (Fig. 2a) [3]. It is currently thought that cbl released from TC inside the lysosome is bound by another putative carrier protein that delivers cbl to a lysosomal transporter (Probable lysosomal cobalamin transporter/Limb region 1 protein homolog (LMBR1) domain-containing protein 1, LMBD1) that releases cbl to the cytosol [21]. Upon export from the lysosome, cobalamins are processed by the cblC gene product MMACHC (methylmalonic aciduria cblC type with homocystinuria) [24, 31, 32] and delivered to cytosolic MS and mitochondrial MMCM by the cblD gene product MMADHC (methylmalonic aciduria cblD type with homocystinuria) [16].

Mutations in the human *LMBRD1* gene that encodes the LMBD1 transporter cause combined methylmalonic aciduria and homocystinuria [49]. Mutations in *LMBRD1* represent one of eight complementation groups of inborn errors of cbl metabolism referred to as cblF. This genetic defect in lysosomal cbl release was discovered 25 years ago, well before the likely transporter involved was characterized [47]. These early studies showed that chloroquine (a compound that increases lysosomal pH above its physiological level of ~ 4.5 thereby inhibiting lysosomal protease activity) prevented the release of cbl from TC and also blocked the transport of lysosomal [57Co] Cbl to both MS and MMCM [47]. These studies provided crucial

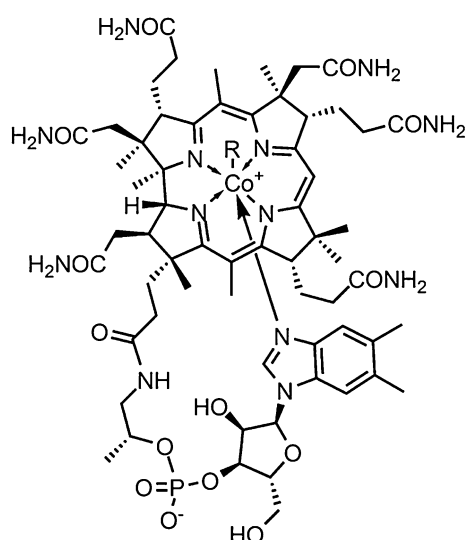


Fig. 1 Cobalamin structure. R = 5'-deoxyadenosyl in 5'-deoxyadenosylcobalamin (AdoCbl), R = CH₃ in methylcobalamin (MeCbl)

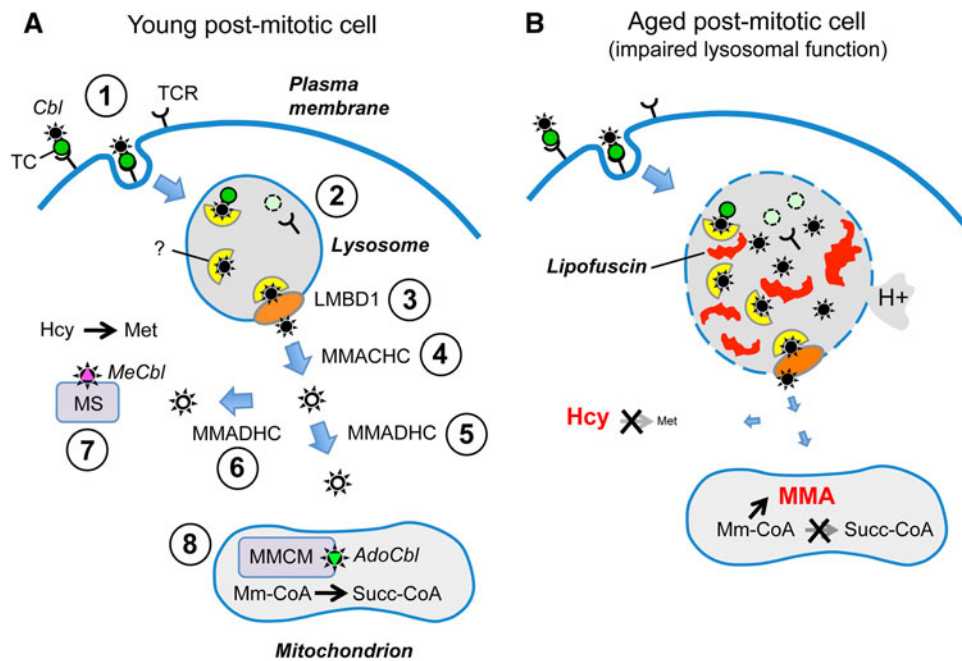


Fig. 2 Lysosomal trafficking of cbl in young and aged post-mitotic cells. **a** TCR-mediated endocytosis delivers cbl to the lysosome (1). Cbl is released from TC in the lysosome (2) and transported by an unknown carrier (?) to LMBD1 for delivery to the cytosol (3). Upon export from the lysosome, cobalamins are processed by MMACHC (4) and delivered via MMADHC (5) and (6) to cytosolic MS (7) and mitochondrial MMCM (8). Cytosolic MeCbl is required for conversion of Hcy to Met by MS (7). Mitochondrial AdoCbl is required for conversion of Mm-CoA to Succ-CoA by MMCM (8). **b** Aging processes (including lipofuscin accumulation) impair lysosomal enzyme function and induce proton “leakage” thereby increasing lysosomal pH. This is predicted to inhibit the release of cbl from TC (due to reduced activity and availability of acid hydrolyases) and

impede the conversion of cbl to the “base-off” state (requires low pH). Overall, this will prevent delivery of cbl to MS and MMCM. As a result, Hcy and Mm-CoA homeostasis is disturbed leading to accumulation of Hcy and the Mm-CoA metabolite MMA that both contribute to cytotoxicity. *Cbl* cobalamin, *TCR* transcobalamin receptor, *TC* transcobalamin, *LMBD1* Probable lysosomal cobalamin transporter, *MeCbl* methylcobalamin, *Hcy* homocysteine, *Met* methionine, *MS* methionine synthase, *AdoCbl* 5'-deoxyadenosylcobalamin, *Mm-CoA* methylmalonyl coenzyme-A, *Succ-CoA* succinyl coenzyme-A, *MMCM* methylmalonyl-CoA mutase, *MMACHC* methylmalonic aciduria cblC type with homocystinuria, *MMADHC* methylmalonic aciduria cblD type with homocystinuria

evidence that lysosomal dysfunction severely impedes intracellular cbl utilization.

In addition to the chloroquine-induced increase in lysosomal pH and concomitant inhibition of protease activity that prevents release of cbl from TC, it is now recognized that the acidic pH of the lysosome also influences the conversion of cbl from the so called “base-on” to “base-off” state that refers to the interaction of the dimethylbenzimidazole moiety of the cbl molecule with the central Co atom [9]. The cbl base-off state is speculated to be important for subsequent interactions with cytosolic cargo proteins [9]. There is strong evidence indicating that decreased lysosomal proteolytic activity and increased lysosomal pH occur as a consequence of aging in long-lived post-mitotic cells [36, 45, 59, 60]. It is therefore plausible that loss of lysosomal TC proteolysis and inhibition of the pH-dependent conversion of cbl to the base-off state accompany such age-related changes in lysosomal function. It is noteworthy that in addition to TC, at least one additional uncharacterized lysosomal cbl escort/transport protein is predicted to exist and this may present

another point at which cbl transit could be disrupted when lysosome function is impaired [10].

Age-related impairment of lysosomal function—exquisite sensitivity of post-mitotic cells

Aging is accompanied by progressive cellular accumulation of macromolecular damage [59]. Although damaged macromolecules and organelles are continuously degraded by lysosomes through autophagy and replaced by newly synthesized biological structures, it is clear that some material progressively accumulates in post-mitotic cells. This is demonstrated by the accumulation of the “age-pigment” lipofuscin in long-lived non-dividing cells such as neurons, cardiac myocytes, skeletal muscle fibers, and retinal pigment epithelial cells [19, 58]. It is well known that cellular lipofuscin content positively correlates with oxidative stress and mitochondrial damage [55]. Furthermore, there is compelling data to suggest that lipofuscin accumulation impairs several lysosomal functions [13, 60].

Lysosomal enzymes are produced in the trans-Golgi network and are transported by secretory vesicles to late endosomes that acidify and mature into lysosomes. The continual fusion and fission of the lysosomal vacuoles ensures the distribution of acid hydrolases within the lysosomal compartment. Senescent post-mitotic cells contain large numbers of lipofuscin-containing lysosomes, to which a progressively greater proportion of lysosomal enzymes are directed in a futile attempt to degrade lipofuscin. These lysosomal enzymes are essentially lost for useful purposes (e.g., for the degradation of newly autophagocytosed material), resulting in a delayed turnover and the accumulation of waste products [59]. There is also evidence that lipofuscin accumulation may contribute to neurodegeneration [70] and that it may exacerbate neuropathological processes that, for example, may be caused primarily by an accumulation of toxic protein aggregates such as huntingtin [11, 56].

Even though lipofuscin-loaded lysosomes appear to be intact using microscopy techniques, it is now recognized that they are subjected to a high Fe-catalyzed oxidative stress [67] that compromises lysosomal membrane integrity leading to loss of the proton gradient [29]. The resulting increase in lysosomal pH significantly reduces protease action and if the lysosomal membrane is sufficiently damaged, cathepsins may be released to the cytosol and trigger apoptosis [68]. Although lysosomal heterogeneity exists both within cells and between cells it is quite clear that the net function of the lysosomal compartment is severely compromised with aging [64].

In addition to the impact of lysosomal lipofuscin accumulation, there are several age-related disorders in which lysosomal function is compromised. Neurodegenerative diseases including Alzheimer's disease, Parkinson's disease (and other Lewy body disorders) and Huntington's disease all involve the accumulation of aggregated proteins/peptides that eventually overwhelm lysosomal capacity for degradation [17, 34, 42, 48]. There is strong evidence that in these conditions the degradative capacity of the lysosome is impaired and the lysosomal membrane is destabilized [42]. In addition, lysosomal pH may be increased in specific lysosomal storage diseases (e.g., mucopolipidosis Type IV), even in dividing cells [5].

Together these data indicate that lysosomal function deteriorates with age and in age-related degenerative disorders. Given the critical role that the lysosome plays in cbl metabolism (described above), it seems reasonable to suggest that cbl probably does not reach its intended intracellular targets in aged/lysosome-compromised cells, even though cbl supply to the cell may be adequate (Fig. 2b). An escalating cytotoxic trajectory would result due to the lack of both MeCbl and AdoCbl available to act

as cofactors in the two important intracellular pathways described below.

Metabolic functions of MeCbl and AdoCbl—relevance to aging and neurodegenerative disease

Cbl has a complex chemical structure with a central cobalt atom tethered equatorially to four nitrogens donated by the corrin ring [27]. Although cbl exists in several forms, MeCbl and AdoCbl are the active forms in human metabolism. MeCbl and AdoCbl differ only in the functional group attached to the Co atom at the centre of the cbl corrin ring (Fig. 1). MeCbl is used to transform Hcy to Met via cytosolic MS. This reaction requires donation of a methyl group from *N*5-methyltetrahydrofolate to Hcy with the formation of tetrahydrofolate and Met. Met is utilized in the formation of *S*-adenosylmethionine (SAM), a universal methyl donor for numerous substrates, including DNA, RNA, hormones, proteins, and lipids. AdoCbl is required for the conversion of methylmalonyl-coenzyme A (Mm-CoA) to succinyl-coenzyme A (Succ-CoA) via mitochondrial MMCM. Succ-CoA then enters the Krebs cycle from where it may be utilized in many pathways including conversion to succinate which may be used as an electron donor or in the synthesis of porphyrins such as heme. In human cbl deficiency states, MS and MMCM activities are reduced which results in increased tissue and plasma Hcy levels and, subsequent to conversion of Mm-CoA to MMA, increased tissue and plasma MMA levels. Based on the roles of both MS and MMCM in methylation reactions and multiple pathways related to, for example, amino acid and lipid metabolism, it is not surprising that clinical cbl deficiency phenotypes are multifaceted [6]. In addition to the “loss of function” caused by impaired MS and MMCM activities, the accumulation of Hcy and MMA is cytotoxic which is thought to contribute to neuron death and to the loss of cognitive capacity [33, 40].

These findings are particularly important in the aging context as it is known that plasma levels of both Hcy and MMA increase with age and there is a positive correlation between plasma Hcy and MMA concentrations and cognitive decline [26, 50, 65]. Importantly, several studies have reported that plasma Hcy levels are positively correlated with brain atrophy in humans and this has led to the administration of cbl (both with and without folate) as a therapeutic agent for mild cognitive impairment (MCI) and Alzheimer's disease [50, 52, 65]. Although there is evidence that reducing Hcy levels in MCI patients can slow the rate of brain atrophy, the response appears to be linked to baseline Hcy levels [54]. Studies in rodents indicate that cbl supplementation significantly improves cognitive

performance [69, 71]. In human MCI and Alzheimer's disease cbl trials, however, the evidence for a positive effect on cognitive performance is not compelling [8, 35, 38, 39, 53].

Although food-Cbl malabsorption is an important cause of cbl deficiency in the elderly, this is not likely to account for the lack of efficacy regarding cognitive improvement in clinical trials since both oral cbl supplementation (that by-passes problems associated with release from food components) and parenteral delivery (that by-passes problems associated with both release from food as well as lack of IF) routes increase circulating cbl to the same degree in both young and aged subjects [4, 41]. This raises the possibility that other pathways that are independent of dietary malabsorption may contribute to suboptimal cbl utilization in aged individuals. One possibility that has not been previously recognized is that the lack of cognitive improvement may be due to impaired transit of cbl through lysosomes within neurons of aged individuals. Such a situation could arise due to the accumulation of lipofuscin in neurons (Fig. 2b), as we have discussed above. In addition, in neurodegenerative diseases including Alzheimer's disease and Parkinson's disease where lysosomal function is impaired [42], it is also likely that cbl release from lysosomal TC is compromised and conversion to the "base-off" state is suboptimal. Under these conditions, a localized cbl deficiency state would prevail despite the fact that plasma cbl levels may have been normalized by dietary supplements or intramuscular injections. Such problems with lysosomal function would help to explain why cbl supplementation has not provided consistent therapeutic benefits that have been hoped for in terms of improved cognitive capacity.

Concluding remarks

In conclusion, the available evidence points towards an impairment of lysosomal function as representing a novel "roadblock" that prevents cbl from reaching its target intracellular enzymes in long-lived post-mitotic cells such as neurons. This may represent a significant cause of "functional cbl deficiency" in aging and neurodegenerative diseases even when oral/parenteral cbl supplementation is used to maintain plasma cbl levels within a healthy range. This roadblock could contribute to the deleterious increases in Hcy and MMA levels that occur in the aging brain and thereby directly accelerate neurodegeneration. As there is already great interest in provision of dietary supplements of cbl and other B-group vitamins in the aging and neurodegenerative disease contexts [7, 23, 53], we feel that future detailed studies of intracellular cbl transport under conditions relevant to aging and impaired lysosomal function are

warranted. Experimental approaches that may be useful in this context include assessment of cbl transport and Hcy/Mm-CoA homeostasis in cell culture models known to induce lysosomal lipofuscin accumulation [13, 44, 57] or impairment of lysosomal function due to pathogenic (e.g., amyloid-beta-induced) lysosomal membrane perturbations [28], as well as in mouse models of ageing, neurodegenerative diseases and certain lysosomal storage disorders [1, 12, 66]. It is conceivable that approaches to preserve lysosomal function or by-pass the dysfunctional lysosome roadblock may be explored in the future as novel strategies to escort cbl to its correct intracellular targets.

References

1. Ahmed Z, Sheng H, Xu YF, Lin WL, Innes AE, Gass J, Yu X, Wuertzer CA, Hou H, Chiba S, Yamanouchi K, Leissring M, Petrucelli L, Nishihara M, Hutton ML, McGowan E, Dickson DW, Lewis J (2010) Accelerated lipofuscinosis and ubiquitination in granulin knockout mice suggest a role for progranulin in successful aging. *Am J Pathol* 177:311–324
2. Allen LH (2009) How common is vitamin B-12 deficiency? *Am J Clin Nutr* 89:693S–696S
3. Amagasaki T, Green R, Jacobsen DW (1990) Expression of transcobalamin II receptors by human leukemia K562 and HL-60 cells. *Blood* 76:1380–1386
4. Andres E, Affenberger S, Vinzio S, Kurtz JE, Noel E, Kaltenbach G, Maloisel F, Schlienger JL, Blickle JF (2005) Food-cobalamin malabsorption in elderly patients: clinical manifestations and treatment. *Am J Med* 118:1154–1159
5. Bach G, Chen CS, Pagano RE (1999) Elevated lysosomal pH in mucopolidosis type IV cells. *Clin Chim Acta* 280:173–179
6. Baik HW, Russell RM (1999) Vitamin B12 deficiency in the elderly. *Annu Rev Nutr* 19:357–377
7. Bailey LB (2004) Folate and vitamin B12 recommended intakes and status in the United States. *Nutr Rev* 62:S14–S20
8. Balk EM, Raman G, Tatsioni A, Chung M, Lau J, Rosenberg IH (2007) Vitamin B6, B12, and folic acid supplementation and cognitive function: a systematic review of randomized trials. *Arch Intern Med* 167:21–30
9. Banerjee R (2006) B12 trafficking in mammals: A for coenzyme escort service. *ACS Chem Biol* 1:149–159
10. Banerjee R, Gherasim C, Padovani D (2009) The tinker, tailor, soldier in intracellular B12 trafficking. *Curr Opin Chem Biol* 13:484–491
11. Bayram-Weston Z, Jones L, Dunnett SB, Brooks SP (2011) Light and electron microscopic characterization of the evolution of cellular pathology in YAC128 Huntington's disease transgenic mice. *Brain Res Bull*. (Epub ahead of print)
12. Bronson RT, Donahue LR, Johnson KR, Tanner A, Lane PW, Faust JR (1998) Neuronal ceroid lipofuscinosis (nclf), a new disorder of the mouse linked to chromosome 9. *Am J Med Genet* 77:289–297
13. Brunk UT, Terman A (2002) Lipofuscin: mechanisms of age-related accumulation and influence on cell function. *Free Radic Biol Med* 33:611–619
14. Calvaresi E, Bryan J (2001) B vitamins, cognition, and aging: a review. *J Gerontol B Psychol Sci Soc Sci* 56:P327–P339
15. Carmel R (1995) Malabsorption of food cobalamin. *Baillieres Clin Haematol* 8:639–655

16. Coelho D, Suormala T, Stucki M, Lerner-Ellis JP, Rosenblatt DS, Newbold RF, Baumgartner MR, Fowler B (2008) Gene identification for the cblD defect of vitamin B12 metabolism. *N Engl J Med* 358:1454–1464
17. Cuervo AM, Stefanis L, Fredenburg R, Lansbury PT, Sulzer D (2004) Impaired degradation of mutant alpha-synuclein by chaperone-mediated autophagy. *Science* 305:1292–1295
18. Dali-Youcef N, Andres E (2009) An update on cobalamin deficiency in adults. *QJM* 102:17–28
19. Double KL, Dedov VN, Fedorow H, Kettle E, Halliday GM, Garner B, Brunk UT (2008) The comparative biology of neuromelanin and lipofuscin in the human brain. *Cell Mol Life Sci* 65:1669–1682
20. Fyfe JC, Madsen M, Hojrup P, Christensen EI, Tanner SM, de la Chapelle A, He Q, Moestrup SK (2004) The functional cobalamin (vitamin B12)-intrinsic factor receptor is a novel complex of cubilin and amnionless. *Blood* 103:1573–1579
21. Gailus S, Hohne W, Gasnier B, Nurnberg P, Fowler B, Rutsch F (2010) Insights into lysosomal cobalamin trafficking: lessons learned from cblF disease. *J Mol Med* 88:459–466
22. Garner B, Roberg K, Brunk UT (1998) Endogenous ferritin protects cells with iron-laden lysosomes against oxidative stress. *Free Radic Res* 29:103–114
23. Gillette Guyonnet S, Abellan Van Kan G, Andrieu S, Barberger Gateau P, Berr C, Bonnefoy M, Dartigues JF, de Groot L, Ferry M, Galan P, Hercberg S, Jeandel C, Morris MC, Nourhashemi F, Payette H, Poulain JP, Portet F, Roussel AM, Ritz P, Rolland Y, Vellas B (2007) IANA task force on nutrition and cognitive decline with aging. *J Nutr Health Aging* 11:132–152
24. Hannibal L, Kim J, Brasch NE, Wang S, Rosenblatt DS, Banerjee R, Jacobsen DW (2009) Processing of alkylcobalamins in mammalian cells: A role for the MMACHC (cblC) gene product. *Mol Genet Metab* 97:260–266
25. He Q, Madsen M, Kilkenney A, Gregory B, Christensen EI, Vorum H, Hojrup P, Schaffer AA, Kirkness EF, Tanner SM, de la Chapelle A, Giger U, Moestrup SK, Fyfe JC (2005) Amnionless function is required for cubilin brush-border expression and intrinsic factor-cobalamin (vitamin B12) absorption in vivo. *Blood* 106:1447–1453
26. Herrmann W, Schorr H, Bodis M, Knapp JP, Muller A, Stein G, Geisel J (2000) Role of homocysteine, cystathionine and methylmalonic acid measurement for diagnosis of vitamin deficiency in high-aged subjects. *Eur J Clin Invest* 30:1083–1089
27. Hodgkin DC, Kamper J, Mackay M, Pickworth J, Trueblood KN, White JG (1956) Structure of vitamin B12. *Nature* 178:64–66
28. Ji ZS, Miranda RD, Newhouse YM, Weisgraber KH, Huang Y, Mahley RW (2002) Apolipoprotein E4 potentiates amyloid beta peptide-induced lysosomal leakage and apoptosis in neuronal cells. *J Biol Chem* 277:21821–21828
29. Kågedal K, Johansson U, Ollinger K (2001) The lysosomal protease cathepsin D mediates apoptosis induced by oxidative stress. *FASEB J* 15:1592–1594
30. Kapadia CR, Donaldson RM Jr (1985) Disorders of cobalamin (vitamin B12) absorption and transport. *Annu Rev Med* 36:93–110
31. Kim J, Gherasim C, Banerjee R (2008) Decyanation of vitamin B12 by a trafficking chaperone. *Proc Natl Acad Sci USA* 105:14551–14554
32. Kim J, Hannibal L, Gherasim C, Jacobsen DW, Banerjee R (2009) A human vitamin B12 trafficking protein uses glutathione transferase activity for processing alkylcobalamins. *J Biol Chem* 284:33418–33424
33. Kolker S, Ahlemeyer B, Kriegelstein J, Hoffmann GF (2000) Methylmalonic acid induces excitotoxic neuronal damage in vitro. *J Inher Metab Dis* 23:355–358
34. Levine B, Kroemer G (2008) Autophagy in the pathogenesis of disease. *Cell* 132:27–42
35. Maron BA, Loscalzo J (2009) The treatment of hyperhomocysteinemia. *Annu Rev Med* 60:39–54
36. Martinez-Vicente M, Sovak G, Cuervo AM (2005) Protein degradation and aging. *Exp Gerontol* 40:622–633
37. McCaddon A (2006) Homocysteine and cognition—a historical perspective. *J Alzheimers Dis* 9:361–380
38. McCaddon A, Hudson PR (2010) L-methylfolate, methylcobalamin, and N-acetylcysteine in the treatment of Alzheimer's disease-related cognitive decline. *CNS Spectr* 15:2–5
39. McMahon JA, Green TJ, Skeaff CM, Knight RG, Mann JI, Williams SM (2006) A controlled trial of homocysteine lowering and cognitive performance. *N Engl J Med* 354:2764–2772
40. Morris MS (2003) Homocysteine and Alzheimer's disease. *Lancet Neurol* 2:425–428
41. Nilsson-Ehle H (1998) Age-related changes in cobalamin (vitamin B12) handling. Implications for therapy. *Drug Aging* 12:277–292
42. Nixon RA, Yang DS, Lee JH (2008) Neurodegenerative lysosomal disorders: a continuum from development to late age. *Autophagy* 4:590–599
43. Quadros EV, Regec AL, Khan KM, Quadros E, Rothenberg SP (1999) Transcobalamin II synthesized in the intestinal villi facilitates transfer of cobalamin to the portal blood. *Am J Physiol* 277:G161–G166
44. Quinn CM, Kågedal K, Terman A, Stroikin U, Brunk UT, Jessup W, Garner B (2004) Induction of fibroblast apolipoprotein E expression during apoptosis, starvation-induced growth arrest and mitosis. *Biochem J* 378:753–761
45. Rajawat YS, Hilioti Z, Bossis I (2009) Aging: central role for autophagy and the lysosomal degradative system. *Ageing Res Rev* 8:199–213
46. Reynolds E (2006) Vitamin B12, folic acid, and the nervous system. *Lancet Neurol* 5:949–960
47. Rosenblatt DS, Hosack A, Matiaszuk NV, Cooper BA, Laframboise R (1985) Defect in vitamin B12 release from lysosomes: newly described inborn error of vitamin B12 metabolism. *Science* 228:1319–1321
48. Rubinsztein DC (2006) The roles of intracellular protein-degradation pathways in neurodegeneration. *Nature* 443:780–786
49. Rutsch F, Gailus S, Miousse IR, Suormala T, Sagne C, Tolia MR, Nurnberg G, Wittkamp T, Buers I, Sharifi A, Stucki M, Becker C, Baumgartner M, Robenek H, Marquardt T, Hohne W, Gasnier B, Rosenblatt DS, Fowler B, Nurnberg P (2009) Identification of a putative lysosomal cobalamin exporter altered in the cblF defect of vitamin B12 metabolism. *Nat Genet* 41:234–239
50. Sachdev PS, Valenzuela M, Wang XL, Looi JC, Brodaty H (2002) Relationship between plasma homocysteine levels and brain atrophy in healthy elderly individuals. *Neurology* 58:1539–1541
51. Seshadri S, Beiser A, Selhub J, Jacques PF, Rosenberg IH, D'Agostino RB, Wilson PW, Wolf PA (2002) Plasma homocysteine as a risk factor for dementia and Alzheimer's disease. *N Engl J Med* 346:476–483
52. Seshadri S, Wolf PA, Beiser AS, Selhub J, Au R, Jacques PF, Yoshita M, Rosenberg IH, D'Agostino RB, DeCarli C (2008) Association of plasma total homocysteine levels with subclinical brain injury: cerebral volumes, white matter hyperintensity, and silent brain infarcts at volumetric magnetic resonance imaging in the Framingham Offspring Study. *Arch Neurol* 65:642–649
53. Smith AD (2008) The worldwide challenge of the dementias: a role for B vitamins and homocysteine? *Food Nutr Bull* 29:S143–S172
54. Smith AD, Smith SM, de Jager CA, Whitbread P, Johnston C, Agacinski G, Oulhaj A, Bradley KM, Jacoby R, Refsum H (2010)

- Homocysteine-lowering by B vitamins slows the rate of accelerated brain atrophy in mild cognitive impairment: a randomized controlled trial. *PLoS ONE* 5:e12244
55. Sohal RS, Brunk UT (1989) Lipofuscin as an indicator of oxidative stress and aging. *Adv Exp Med Biol* 266:17–26
 56. Stack EC, Matson WR, Ferrante RJ (2008) Evidence of oxidant damage in Huntington's disease: translational strategies using antioxidants. *Ann NY Acad Sci* 1147:79–92
 57. Terman A, Brunk UT (1998) Ceroid/lipofuscin formation in cultured human fibroblasts: the role of oxidative stress and lysosomal proteolysis. *Mech Ageing Dev* 104:277–291
 58. Terman A, Brunk UT (1998) Lipofuscin: mechanisms of formation and increase with age. *APMIS* 106:265–276
 59. Terman A, Brunk UT (2006) Oxidative stress, accumulation of biological 'garbage', and aging. *Antioxid Redox Signal* 8:197–204
 60. Terman A, Gustafsson B, Brunk UT (2006) The lysosomal-mitochondrial axis theory of postmitotic aging and cell death. *Chem Biol Interact* 163:29–37
 61. Terman A, Kurz T, Navratil M, Arriaga EA, Brunk UT (2010) Mitochondrial turnover and aging of long-lived postmitotic cells: the mitochondrial-lysosomal axis theory of aging. *Antioxid Redox Signal* 12:503–535
 62. Torres I, Smith WT, Oxnard CE (1971) Peripheral neuropathy associated with vitamin-B 12 deficiencies in captive monkeys. *J Pathol* 105:125–146
 63. Van den Berg MP, Merkus P, Romeijn SG, Verhoef JC, Merkus FW (2003) Hydroxocobalamin uptake into the cerebrospinal fluid after nasal and intravenous delivery in rats and humans. *J Drug Target* 11:325–331
 64. von Zglinicki T, Nilsson E, Docke WD, Brunk UT (1995) Lipofuscin accumulation and ageing of fibroblasts. *Gerontology* 41(Suppl 2):95–108
 65. Williams JH, Pereira EA, Budge MM, Bradley KM (2002) Minimal hippocampal width relates to plasma homocysteine in community-dwelling older people. *Age Ageing* 31:440–444
 66. Yu WH, Cuervo AM, Kumar A, Peterhoff CM, Schmidt SD, Lee JH, Mohan PS, Mercken M, Farmery MR, Tjernberg LO, Jiang Y, Duff K, Uchiyama Y, Naslund J, Mathews PM, Cataldo AM, Nixon RA (2005) Macroautophagy—a novel beta-amyloid peptide-generating pathway activated in Alzheimer's disease. *J Cell Biol* 171:87–98
 67. Yu Z, Persson HL, Eaton JW, Brunk UT (2003) Intralysosomal iron: a major determinant of oxidant-induced cell death. *Free Radic Biol Med* 34:1243–1252
 68. Yuan XM, Li W, Dalen H, Lotem J, Kama R, Sachs L, Brunk UT (2002) Lysosomal destabilization in p53-induced apoptosis. *Proc Natl Acad Sci USA* 99:6286–6291
 69. Zhang CE, Wei W, Liu YH, Peng JH, Tian Q, Liu GP, Zhang Y, Wang JZ (2009) Hyperhomocysteinemia increases beta-amyloid by enhancing expression of gamma-secretase and phosphorylation of amyloid precursor protein in rat brain. *Am J Pathol* 174:1481–1491
 70. Zhao L, Spassieva SD, Jucius TJ, Shultz LD, Shick HE, Macklin WB, Hannun YA, Obeid LM, Ackerman SL (2011) A deficiency of ceramide biosynthesis causes cerebellar purkinje cell neurodegeneration and lipofuscin accumulation. *PLoS Genet* 7:e1002063
 71. Zhuo JM, Pratico D (2010) Acceleration of brain amyloidosis in an Alzheimer's disease mouse model by a folate, vitamin B6 and B12-deficient diet. *Exp Gerontol* 45:195–201